

RCN: Enzymes in the Environment

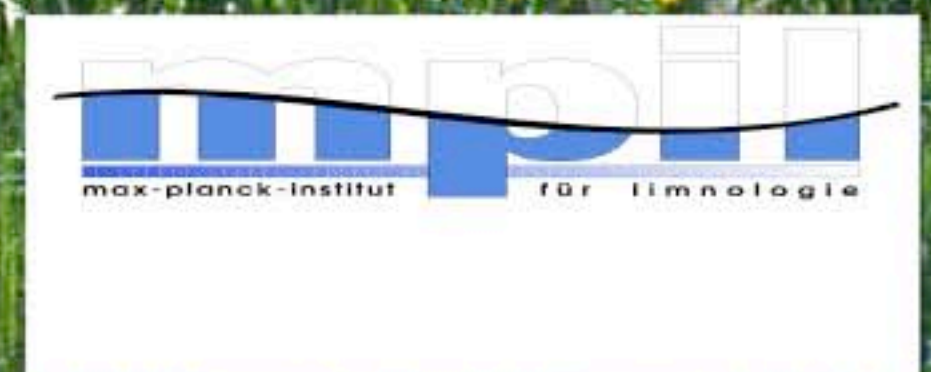
Big questions and challenges: Linking aquatic and terrestrial environments

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Soil and water habitats

Macrohabitats

- characterized by varying proportions of 2 phases: solid surfaces and water
- continuum (?) of habitats: from arid deserts (no or little pore water) to offshore sea water (only traces of particulate matter, lacking nearly completely any solid surfaces)

Fundamental differences between soil and water

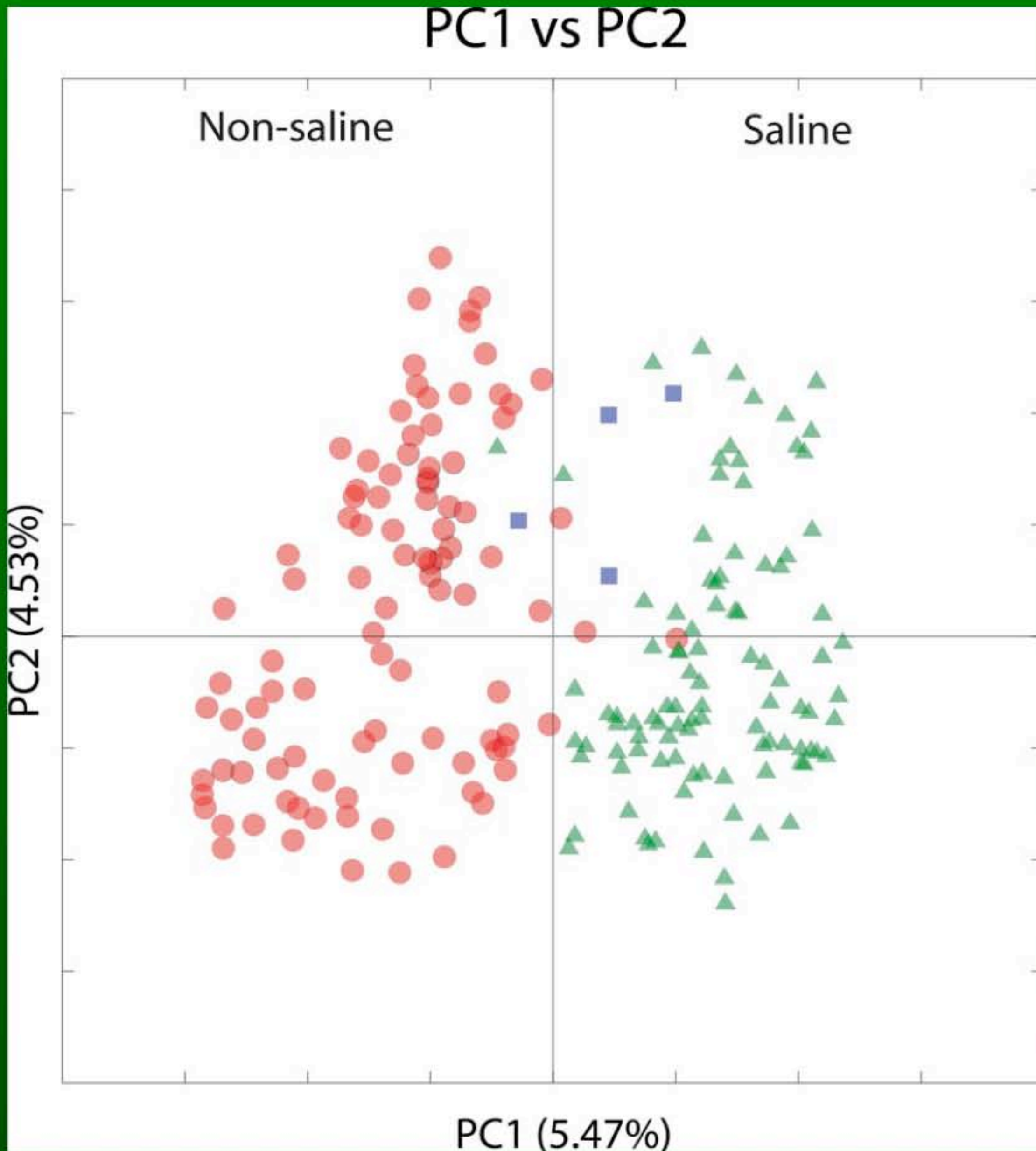
- water availability
- substrates for microbial growth
 - soil: plant residues
 - water: more direct coupling to primary producers
- constancy: more pronounced constancy in aquatic environm.

Microhabitats

are the effective habitats for microorganisms

- aquatic microhabitats occur also in soil

Global patterns in bacterial community composition: salinity

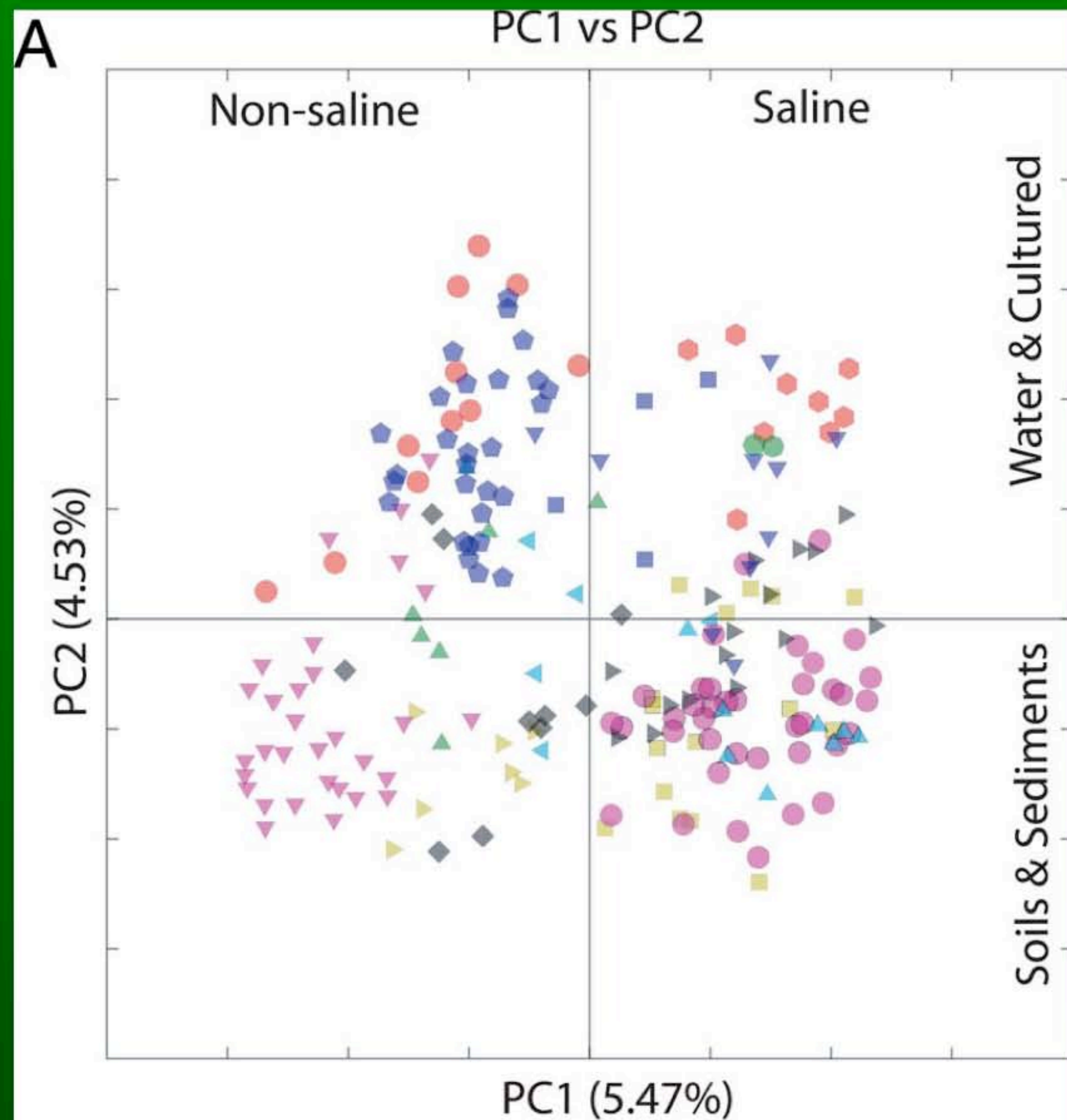


Major environmental determinant: salinity (rather than temperature, pH, or other physical and chemical factors)

red circles – non-saline environments
green triangles – saline environments
blue squares – mixed environments

From LOZUPONE, C.A., & R. KNIGHT, 2007. Global patterns in bacterial diversity. PNAS 104: 11436-11440.

Global patterns in bacterial community composition: water vs. sediment/soil



2nd most important determinant:
substratum type (water vs. soil/
sediment)

blue pentagons – nonsaline water

red circles – nonsaline cultured

purple triangles – surface soils

yellow sideways triangles – sediments

gray diamonds – submerged soils and
aquifers

blue sideways triangles – nonsaline springs

green triangles – nonsaline endolithic

From LOZUPONE, C.A., & R. KNIGHT, 2007. Global
patterns in bacterial diversity. PNAS 104: 11436-11440.

Diversity in soil and aquatic sediments

- Aquatic sediments

- highest phylogenetic diversity from all environments considered (highly stratified nature, chemical gradients)

- Soil

- low phylogenetic diversity, but
- very high species level diversity (more closely related species in one sample)
- fungi not considered

Levels of analyzing structure and function of extracellular enzymes

- Genomics (DNA)
- Transcriptomics (mRNA)
- Proteomics (proteins)
- Metabolomics (metabolites)

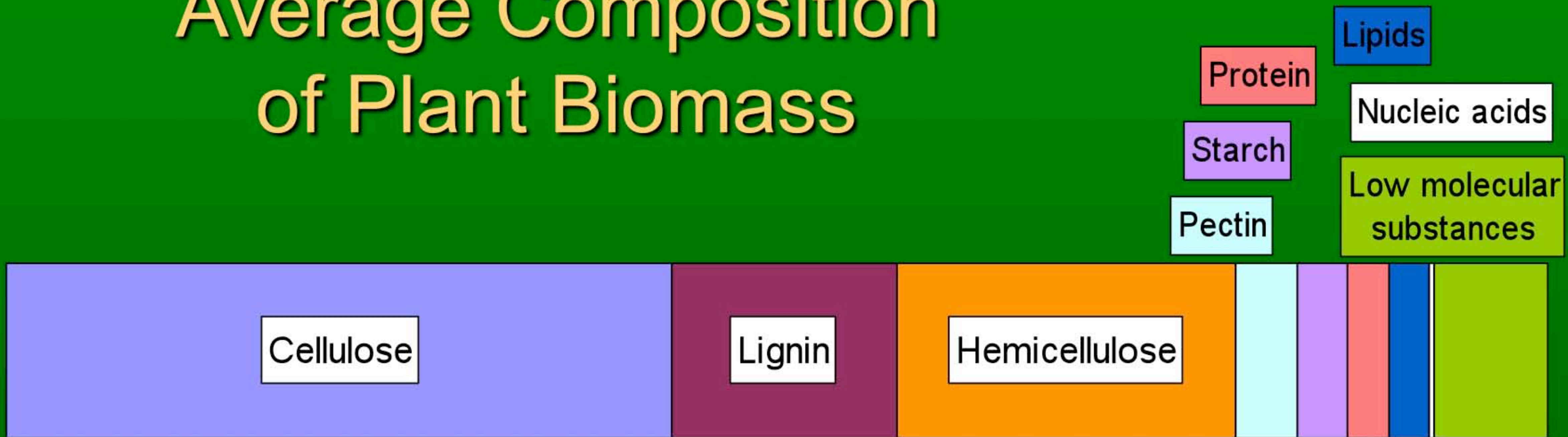
E.g.: RAES, J., & P. BORK, 2008. Molecular eco-systems biology: towards an understanding of community function. Nature Rev. Microbiol. 6: 693-699.

Broad and narrow functions *sensu* Schimel

- Broad processes
 - carried out by many organisms
 - little apparent influence of microbial community structure
- Narrow processes
 - carried out by restricted groups of organisms
 - potential for substantial ecosystem-level effects from differences in microbial diversity and community structure
 - example: litter degradation

SCHIMEL, J., 1995. Ecosystem consequences of microbial diversity and community structure. In: F.S. CHAPIN & C. KÖRNER (eds.), Arctic and Alpine Biodiversity: Patterns, Causes and Ecosystem Consequences. Ecosystem Studies 113, Springer, Berlin, p. 239-254.

Average Composition of Plant Biomass



↓
endoglucanase,
cellobiohydrolase,
 β -glucosidase,
.....

↓
phenol-oxidase,
peroxidase,
.....

↓
 β -xylosidase,
.....

Extracellular enzymes involved in degradation

Narrow functions among extracellular enzymes?

- Are among extracellular enzymes (esp. those involved in the degradation of refractory macromolecules) really narrow functions?
- Are the communities and functions in soil and aquatic environments sensitive to environmental changes?
- Does reduction of community diversity and/or do changes in community composition have large-scale effects on these processes?
- Is there any influence on the overall CO₂ release by soil and aquatic environments?

Water availability

- Soil
 - narrow processes more sensitive to be affected in soil
- Aquatic environments
 - exposed to drought in temporary waters
 - headwaters become more affected by more frequent and longer lasting droughts because of climate change in some regions
 - extracellular enzymes have an important role for recovery of microbial communities and re-establishing its functions after rewetting

MARXSEN, J., A. ZOPPINI & S. WILCZEK, 2010. Microbial communities in streambed sediments recovering from desiccation. FEMS Microbiol. Ecol. 71: 374-386

Possible approach

- Initial metagenome and metaproteome analysis
(+ metatranscriptome analysis?)
 - select functional genes expressed at high enough levels
 - taxonomic analysis can be included
- Detailed analysis of functional genes
 - PCR-based assays (DNA, cDNA)
 - after having selected or developed suitable primers
- Data on the diversity of enzymes
 - genome, transcriptome and protein level
 - compare with activity measurements

Important questions

- How are community composition and extracellular enzyme activity linked?
- Which are the differences between typical aquatic and terrestrial environments regarding enzyme diversity and activity?
- Are there regional differences?
- What about seasonal fluctuations?
- Which is the influence of natural and anthropogenic stressors (e.g. water availability, pH, temperature, nutrient enrichment, contamination)?
-?